Severe Acute Respiratory Syndrome Coronavirus Papain-like Novel Protease Inhibitors: Design, Synthesis, Protein–Ligand X-ray Structure and Biological Evaluation[†]

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The design, synthesis, X-ray crystal structure, molecular modeling, and biological evaluation of a series of new generation SARS-CoV PLpro inhibitors are described. A new lead compound **3** (6577871) was identified via high-throughput screening of a diverse chemical library. Subsequently, we carried out lead optimization and structure—activity studies to provide a series of improved inhibitors that show potent PLpro inhibitor **15h** (enzyme IC₅₀ = 0.56μ M; antiviral EC₅₀ = 9.1μ M) and the corresponding (*R*)-Me **15g** (IC₅₀ = 0.32μ M; antiviral EC₅₀ = 9.1μ M) are the most potent compounds in this series, with nearly equivalent enzymatic inhibition and antiviral activity. A protein—ligand X-ray structure of **15g**-bound SARS-CoV PLpro and a corresponding model of **15h** docked to PLpro provide intriguing molecular insight into the ligand-binding site interactions.

Introduction

Severe acute respiratory syndrome (SARS^{*a*}) was first reported in Guangdong province, China, in November 2002.¹ SARS is a contagious respiratory illness with no effective treatment to date. SARS affected three continents, infecting more than 8000 individuals and causing nearly 800 deaths. Fortunately, the spread of SARS-CoV was contained after the initial outbreaks through public health measures. As it turned out, the etiological agent of SARS is a novel coronavirus, SARS-CoV.^{2,3} There have been no known new cases of SARS since 2005. However, recent isolation of strains from zoonotic origins thought to be the reservoir for SARS-CoV raises the possibility of a reemergence of SARS and related ailments.^{4,5} Consequently, design and development of antivirals effective against SARS-CoV should be an important priority against future outbreaks.

Biochemical events critical to the viral replication revealed a number of important targets for therapeutic intervention of SARS.^{6,7} Most notably, two cysteine proteases, a papain-like protease (PLpro) and a 3C-like protease (3CLpro), play a critical role in the virus-mediated RNA replication. Not surprisingly, numerous studies related to the development of SARS-CoV 3CLpro inhibitors have already been reported.^{8,9} In contrast, very few inhibitor design efforts against SARS-CoV PLpro have been reported. We recently reported the

discovery and design of a series of unprecedented noncovalent SARS-CoV PLpro inhibitors displaying antiviral activity against SARS-CoV with no associated cytotoxicity.¹⁰ Subsequently, a protein–ligand X-ray structure provided important molecular insights for further design and optimization of inhibitors.¹⁰ This initial work demonstrated that PLpro is a viable target for the development of anti-SARS therapeutics.

Besides viral peptide cleavage, recent structural and functional studies demonstrated that PLpro is involved in a number of other important biochemical events, such as deubiquitination, deISGylation, and involvement in the virus evasion from the innate immune response.^{11,12} The homologous enzyme PLP2, from the human coronavirus 229E, has been shown to be critical to 229E viral replication.¹³ In addition, recent studies have shown that human deubiquitinating enzymes are potential anticancer drug-design targets. Thus, PLpro is a significant target for development of drugs against SARS and is a model for development of drugs against other deubiquitinating enzymes involved in human diseases.

Recently, our primary screening of a library of 50 080 diverse, druglike compounds led to the identification of two compounds after lead validation. Both leads reproducibly inhibited PLpro in a dose dependent manner in the absence and presence of Triton-X. Subsequently, our optimization efforts of the most potent lead, **1** (7724772), containing a benzamide scaffold (IC₅₀ = 20.1 ± 1.1 μ M) led to the design of novel PLpro inhibitor **2** and related derivatives that displayed antiviral activity against SARS-CoV. We recently reported a detailed study describing synthesis, biological studies, and X-ray structure of the protein–ligand complex of **2**-bound PLpro.¹⁰ In our continuing studies toward the development of noncovalent/reversible PLpro inhibitors, we

 $^{^{\}dagger}\text{The PDB}$ accession code for 15g-bound PLpro X-ray structure is 3MJ5.

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^{*a*} Abbreviations: SARS, severe acute respiratory syndrome; SARS-CoV, severe acute respiratory syndrome coronavirus; 3CLpro, chymo-trypsin-like protease; PLpro, papain-like protease; WHO, World Health Organization.



Figure 1. Structures of PLpro inhibitors 1–3 and 15g.

have now investigated the potential of the second and less potent lead that evolved from our high-throughput screening efforts. The second HTS lead, compound 3 (Figure 1), contains a piperidine carboxamide scaffold and exhibited an IC₅₀ value of 59 μ M. Our subsequent lead optimization efforts led to the design of potent inhibitor 15g (IC₅₀ = 0.32 μ M) which inhibited SARS-CoV viral replication in Vero cells with an EC_{50} value of 9.1 μ M. The corresponding enantiomer **15h** has shown slightly less potent enzyme inhibitory activity (IC₅₀ = 0.56 μ M) and similar antiviral potency. A protein-ligand X-ray structure of 15g-bound SARS-CoV PLpro was determined. Interestingly, this structure revealed a unique mode of binding with SARS-CoV PLpro and that key molecular interactions of inhibitor 15g are quite different from the active-site interactions with inhibitor 2. Herein we describe the design, synthesis, structure-activity studies, molecular modeling, protein-ligand X-ray structure, and biological evaluation of a series of novel and noncovalent inhibitors of SARS-CoV PLpro.

Chemistry

To ascertain the importance of the position of the methoxy substituent in lead inhibitor **3**, we have synthesized the corresponding 2-methoxy and 3-methoxybenzyl derivatives. As shown in Scheme 1, Boc-piperidine-4-carboxylic acid **4** was coupled with 2- and 3-methoxybenzylamines **5a** and **5b** using *N*-(3-dimethylaminopropyl)-*N'*-ethylcarbodiimide hydrochloride (EDCI) and 1-hydroxybenzotriazole hydrate (HOBT) in the presence of *N*-methylmorpholine (NMM) in CH₂Cl₂ to provide coupling products **6a** and **6b** in 92% and 94% yield, respectively. Removal of Boc-group by exposure to trifluoroacetic acid (TFA) in CH₂Cl₂ at 0–23 °C for 6 h afforded the respective amine. Reductive amination of these amines with 1-naphthaldehyde using Na(OAc)₃BH in the presence of acetic acid furnished inhibitors **7a** and **7b** in 70% and 71% yield, respectively.

Scheme 1^a



^{*a*}Reagents and conditions: (a) **5a** or **5b**, EDCI, HOBT, NMM, CH₂Cl₂, 23 °C, 5 h; (b) TFA, 0-23 °C, 6 h; (c) 1-naphthaldehyde, Na(OAc)₃BH, AcOH, CH₂Cl₂, 23 °C, 12 h.

Scheme 2^{*a*}



^{*a*} Reagents and conditions: (a) KO^{*t*}Bu, DMSO, 23 °C, 48 h; (b) 10% HCl, THF, 23 °C, 18 h; (c) NaHCO₃, 23 °C, 16 h; (c) H₂, PtO₂, EtOAc, 23 °C, 2 h.

For structure–activity studies and optimization of potency, we planned to synthesize derivatives of both 1- and 2-naphthylethylpiperidin-4-carboxylic acids and coupled them with various substituted benzylamine derivatives. The synthesis of substituted piperidine-4-carboxylic acids is shown in Scheme 2. Alkylation of dimethyl malonate **8** with commercially available 2-bromomethyl-1,3-dioxolane **9** in the presence of KO'Bu in DMSO at 23 °C afforded malonate derivative **10** as described previously.¹⁴ Deprotection of the ketal functionalities was carried out by treatment of **10** with 10% aqueous HCl in THF at 23 °C. The reaction was quenched with solid NaHCO₃, and the resulting crude dialdehyde was used directly for the subsequent condensation reaction. Condensation of the dialdehyde with various

Scheme 3^{*a*}



^{*a*} Reagents and conditions: (a) NaCN, DMF, reflux, 16 h; (b) LiOH \cdot H₂O, THF/MeOH/H₂O (3:1:1), 23 °C, 16 h; (c) **5a-d**, EDCI, HOBT, DIPEA, CH₂Cl₂/DMF (9:1), 23 °C, 15 h.

Scheme 4^a



^{*a*} Reagents and conditions: (a) NaBH₃CN, MeOH/AcOH (50:1), 23 °C, 48 h; (b) TFA, CH₂Cl₂, 23 °C, 2 h; (c) **5c**, *N*,*N*'-carbonyldiimidazole, CH₂Cl₂, 23 °C, 4 h.

optically active (*S*)- and (*R*)-1-methyl-1-naphthylmethylamines, 1-methyl-2-naphthylmethylamines, 2-naphthylmethylamine, 1-naphthylmethylamine, and dimethyl-1-naphthylmethylamine **11a-g**¹⁰ in aqueous THF for 16 h afforded dihydropyridines **12a**-g in 39–62% yield.^{15,16} Catalytic hydrogenation of dihydropyridines **12a**-f in ethylacetate at 23 °C provided various piperidine derivatives **13a**-f in 60–94% yield.

The synthesis of various test inhibitors is shown in Scheme 3. Treatment of diesters 13a-f with NaCN in DMF at reflux for 16 h provided methyl esters 14a-f in 38-92% yield. Dihydropyridine derivative 12g was similarly converted to methyl ester 14g in a two-step sequence. Saponification of 14a-g with aqueous LiOH in a mixture (3:1:1) of THF, methanol, and water at 23 °C for 16 h afforded the corresponding carboxylic acids. Coupling of these resulting carboxylic acids with benzylamine derivatives 5a-d utilizing EDCI in the presence of diisopropylethylamine as described above furnished various inhibitors 15a-k in excellent yield (80-99%).

To evaluate the effect of the corresponding piperazine derivatives, we sought to synthesize racemic piperazine derivative **20**, and the synthesis is outlined in Scheme 4. Reductive amination¹⁷ of Boc-piperazine **16**¹⁸ with 1-acetonaphthone **17** using sodium cyanoborohydride in a mixture (50:1) of methanol and acetic acid at 23 °C for 48 h afforded **18** in 24% yield. Removal of the Boc-group by treatment with trifluoroacetic acid in CH₂Cl₂ at 23 °C for 2 h provided amine **19**.¹⁹ Treatment

 Table 1. Structure and Activity of 1- and 2-Naphthylmethyl Derivatives^a

Compound	Structure	IC ₅₀ (µM)
3	O Me	59.2 ± 7.8
7a	H O O Me	116 ± 30
7ь	H COMe	30 ± 3
15a		1.21 ± 0.04
15b	N H OME	0.34 ± 0.01
15c	C C C C C C C C C C C C C C C C C C C	0.34 ± 0.01
15d		13.2 ± 0.6
15e		34.8 ± 4.0
15f		5.8 ± 0.1
20	C A C A C A C A C A C A C A C A C A C A	>100

 a NA = not active.

of 4-methoxybenzylamine **5c** in the presence of N,N'-carbonyldiimidazole in CH₂Cl₂ followed by addition of **19** and stirring of the resulting mixture at 23 °C for 4 h afforded piperazine derivative **20** in 90% yield.

Results and Discussion

The second HTS lead **3** is considerably weaker than the first lead inhibitor **1**, a benzamide derivative of 2-naphthylethylamine. To enhance activity, we first investigated the effect of 2-methoxy and 3-methoxy derivatives **7a** and **7b** on PLpro inhibitory activity. As shown in Table 1, 2-methoxy derivative **7a** showed a very poor inhibitory activity. The 3-methoxy

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derivative, 7b, however, displayed slightly better activity than the starting lead 3. Our previous structure-activity of lead 1 established that 1-naphthylethylamides were significantly more potent than the corresponding 2-naphthyl derivative. The X-ray structure of 2 bound to PLpro demonstrated that a (R)-1-naphthylethylamide forms hydrophobic interactions with the Tyr-265 and Tyr-269 aromatic rings and with side chains of Pro-248 and Pro-249.¹⁰ The preference for (R)methyl was also documented, as it points into the interior of the enzyme between Tyr-265 and Thr-302. On the basis of this ligand-binding site interaction, we elected to incorporate the (R)-methyl group. As shown in Table 1, the (R)-methyl derivative 15a displayed an IC₅₀ value of $1.2 \,\mu$ M. To ascertain the importance of the position of the methoxy group, we synthesized *m*-methoxy and *p*-methoxy derivatives. Interestingly, *m*-methoxy derivative **15b** exhibited improvement of enzyme inhibitory activity with an IC₅₀ value of $0.34 \,\mu$ M. The corresponding *p*-methoxy derivative 15c has also shown similar potency enhancement (>170-fold over 3). However, the 2-methoxy derivative 15a showed a 3-fold reduction in potency over 15b and 15c. We then examined the effect of 2-(R)-naphthylethyl derivatives on potency. As shown, both *m*-methoxybenzylamide **15d** and *o*-methoxybenzylamide **15e** displayed significant reductions in potency compared to the 1-(R)-naphthylethyl derivatives 15b and 15a, respectively. Interestingly, the 2-(S)-naphthylethyl derivative 15f is 2-fold more potent than the 2(R)-derivative 15d.

We next examined the effect of a piperazine ring in place of piperidine in **15c** by preparing compound **20**. However, this piperazine derivative showed no activity against PLpro. Most likely, the piperazine derivative showed no activity against PLpro because of the structural constraints imposed by the carbon to nitrogen replacement on this ring. The new nitrogen is then attached to the amide group, forming a urea moiety. This urea moiety will tend to be planar, imposing a flexibility constraint. GOLD docking shows the amide to rotate ~90° away from the optimal hydrogen-bonding orientation (data not shown) of the other active compounds described here.

Our structure-activity studies established that both mmethoxy and *p*-methoxy derivatives (15b and 15c) are equally potent. Our preliminary modeling studies indicated that either methoxy oxygen (meta or para) is within proximity to form a hydrogen bond with the Gln-270 carboxamide side chain. On the basis of these possible interactions, we incorporated a benzodioxolane ring and examined its effect on inhibitory potency. As shown in Table 2, dioxolane derivative 15g exhibits potency comparable to the corresponding m- and *p*-derivatives **15b** and **15c**. The corresponding (S)-derivative 15h also shows comparable enzyme inhibitory activity. To examine the preference for a methyl group over a hydrogen at the 1- and 2-naphthylmethyl positions, we have synthesized and evaluated the corresponding unsubstituted derivatives 15i and 15j. As shown, both compounds displayed significant reduction in potency, indicating the importance of the methyl group. We have also examined the corresponding gemdimethyl derivative 15k. Interestingly, this compound is inactive, indicating that both methyl groups cannot be accommodated by the PLpro active site.

Antiviral activities of selected PLpro inhibitors were determined, and the results are shown in Table 3. The compounds were assayed for their ability to rescue a Vero cell culture from SARS-CoV infection. The viability of virus-infected Vero E6 cells as a function of inhibitor concentration was measured relative to mock-infected cells using a luminescence assay.

Table 2. Structure and Activity of Benzodioxolane Derivartives



 Table 3.
 Evaluation of Compounds as Inhibitors of SARS-CoV Replication in a Cell-Based Assay

compd	IC ₅₀ (µM)	EC ₅₀ (µM)
3	59.2 ± 7.8	NI
15a	1.21 ± 0.04	11.6 ± 0.6
15b	0.34 ± 0.01	9.7 ± 0.3
15c	0.34 ± 0.01	10.2 ± 0.5
15f	5.8 ± 0.1	> 25
15g	0.32 ± 0.01	9.1 ± 0.5
15h	0.56 ± 0.03	9.1 ± 0.3

This protocol allows for the evaluation of both inhibitor efficacy and cytotoxicity. As can be observed from the data presented in Table 3, the original HTS lead (3) does not show any antiviral activity. However, all 2-, 3-, and 4-methoxy derivatives 15a-c show comparable antiviral activity. Inhibitor 15f with a 2-naphthyl substituent displayed no antiviral activity. While the (*R*)-methyl derivative 15g showed slightly better enzyme activity than the (*S*)-methyl derivative 15h, both inhibitors exhibited the same antiviral potency (EC₅₀ = $9.1 \,\mu$ M). Interestingly, both dioxolane derivatives 15g and 15h showed antiviral activity approximately comparable to the activity of the corresponding methoxy or benzamide derivatives reported in our previous studies.¹⁰

To obtain molecular insight into the ligand-binding site interactions, the X-ray crystal structure of **15g** bound to PLpro was determined. Interestingly, the binding mode and key molecular interactions of inhibitor **15g** are quite different than predicted and are different from the active-site interactions with the benzamide-derived inhibitor binds to a loop adjacent to the active site via a series of interactions including a hydrogen-bond formed between the carboxamide NH of the inhibitor and the backbone carbonyl of Tyr-269, with **15g** wrapped around the β -turn. The **15g** bound PLpro crystal structure also confirms the presence of a few structural water molecules conserved between the apo enzyme (PDB code



Figure 2. Stereorepresentation of 15g bound to PLpro, including the conserved waters adjacent to the binding site that may influence the binding conformation, as described in the text.



Figure 3. X-ray structure of inhibitor 15g-bound (yellow) PLpro (gray) (PDB code 3MJ5) superimposed on the X-ray structure of inhibitor 2-bound (cyan) PLpro (pink) (PDB code 3E9S).

2FE8) and inhibitor **2** bound PLpro (PDB code 3E9S). One of the conserved water molecules sits in the P5 pocket shown in Figure 2 as spheres between residues Asp-165, Asp-303, and Thr-302, preventing the inhibitor naphthyl rings from occupying this pocket. In the stereoimage of **15g**-bound PLpro we also show two other water molecules near residue Leu-163 and Lys-158 that may prevent the benzodioxolane ring from flipping down toward Lys-158.

Figure 3 superimposes 15g and our previously developed inhibitor, 2,¹⁰ and demonstrates that the binding mode differs

significantly between the two inhibitors. Interestingly, the turn region between Tyr-269 and Gln-270 also shows significant flexibility, particularly in the case of inhibitor **2** (PDB code 3E9S), where the peptide bond between Tyr-269 and Gln-270 flips by 180° to enable a hydrogen bond interaction between the backbone nitrogen of Tyr-269 and the carboxamide oxygen in inhibitor **2**. The carboxyamide nitrogen makes a hydrogen bond with the side chain carboxylate of Asp-165. The carboxy amide nitrogen of inhibitor **15g** (yellow) forms a hydrogen bond with the backbone carbonyl



Figure 4. (A) Superposition of enantiomer 15h (blue) with the crystal structure of 15g-bound (yellow) PLpro. (B) Docked alignment of the *gem*-dimethyl substituted compound in the 15g-ligand removed PLpro crystal structure. The bumping collision of one of the methyl groups of the *gem*-dimethyl (magenta) 15k with the Asp-165 carboxylate is noted.

oxygen of Tyr-269 (protein shown in gray). The naphthyl rings of both inhibitors **2** and **15g** align in a similar fashion in the hydrophobic pocket formed by residues Tyr-269, Tyr-265, Pro-248, Pro-249, and Thr-302. The overlapping position of one conserved water molecule observed for both the inhibitor **2**-bound PLpro (oxygen atom shown as sphere in pink) crystal structure and inhibitor **15g**-bound PLpro (oxygen atom shown as sphere in red) crystal structure is shown as overlapping spheres.

Modeling Studies

To understand the SAR of the analogues of HTS hit compound 3, we used computer modeling to explore the interactions of this series of inhibitors with PLpro. The activity of this series of compounds is independent of stereoisomerism in contrast to the series of compounds synthesized from the first HTS hit compound 1.¹⁰ GOLD redocking of inhibitor 15g into the PLpro crystal structure described above produces a heavy atom rmsd of 1.7 Å with the crystal structure conformation of 15g, indicating that docking satisfactorily reproduces the experimental structure. When the inhibitors 15g, 15h, and 15k are docked into the ligand removed 15g-bound PLpro crystal structure (with residues Tyr-269 and Gln-270 flagged as flexible), the internal strain scores of the compounds correlate very well with their enzymatic activities. The conserved overlapping water molecules observed in both chains A and B of the 15g-bound PLpro crystal structure were included for all docking studies.

To investigate the structural basis of the potency insensitivity to the (R)-Me (15g) versus (S)-Me (15h) configuration, we show the docked model of inhibitor 15h superimposed on the crystal structure of 15g-bound PLpro in Figure 4A. From this model, we observe an inversion of the piperidine ring between the (R)-Me and (S)-Me binding modes that allows the naphthyl rings of both isomers to be accommodated in the active site in very similar orientations. The flexible piperidine ring also acts as a spacer group that enables the carboxamide NH of both 15h and 15g to hydrogen-bond with the backbone carbonyl oxygen of Tyr-269 in a similar fashion, thereby retaining the potency of both enantiomers. However, the *gem*dimethyl substitution in 15k decreases the freedom around the carbon atom and locks the compound in a conformation where one of the methyl groups exhibits a bumping collision with the side chain of Asp-165. One of the methyl groups in **15k** shifts almost 1.2 Å toward residue Asp-165 when compared to the single methyl substitution (R)-Me in **15g**, as can be seen in Figure 4B. It is important to note that the side chain of this Asp-165 is locked in its position by a hydrogen bond with the backbone NH of Arg-167. Hence, the *gem*-dimethyl substitution is not favorably accommodated in the active site because in order to fit the hydrophobic methyl group near the hydrophilic residue, the aspartic acid side chain would have to move out, thereby breaking structural hydrogen bonding with Arg-167.

This hypothesis is further validated by the GoldScore scoring function of GOLD, version 4.1, during the docking study. Compound 15k is heavily penalized because of an unfavorable internal energy term (-12 compared to about)-6 for both 15g and 15h) which is a sum of the internal torsional strain and internal van der Waals energy terms of the ligand. Docking with flexible residues also suggests that the Gln-270 side chain may adopt conformations that might enable hydrogen bonding interactions with one of the 1,3 benzodioxolane oxygens in 15g and 15h (within 3 Å). However, all docked conformations generated for 15k show a loss of this hydrogen bonding interaction. The closest benzodioxolane oxygen of 15k is at least 4.8 Å away from the side chain of Gln-270 (not shown). Figure 4B highlights the potential bumping collision of one of the methyl groups of 15k with Asp-165, demonstrating that two methyl groups cannot be accommodated favorably at this position.

In our previous study, we discussed the SAR of the analogues of our first HTS hit 1 and the evolution of inhibitor 2 in great detail.¹⁰ In distinct contrast to the present work, that series of compounds is extremely sensitive to the enantiomeric form of the compound. From docking studies we concluded that the (R)-Me form was active whereas the (S)-Me was inactive because the (S)-Me conformation pushed the carbox-amide group of the inhibitor away from the backbone NH of Tyr-269, inhibiting hydrogen bond formation with the loop residue.

Conclusion

We have designed, synthesized, and evaluated a novel series of SARS-CoV PLpro inhibitors. Initial lead structure **3** $(IC_{50} = 59.2 \,\mu\text{M})$ was discovered via high-throughput screening of a library of diverse compounds. Our preliminary structure-activity studies and systematic modification guided by X-ray crystal structure of 2-bound PLpro and subsequent molecular modeling resulted in a potent inhibitor 15g with enzyme inhibitory IC₅₀ value of 320 nM and antiviral EC₅₀ value of 9.1 µM in SARS-CoV-infected Vero E6 cells. Interestingly, the corresponding (S)-isomer **15h** is only slightly less potent (IC₅₀ = 560 nM) in PLpro inhibitory assays but equipotent in antiviral assays. The corresponding gem-dimethyl derivative 15k is significantly less potent. A protein-ligand X-ray structure of 15g-bound PLpro was determined to 2.6 Å resolution. This structure provided critical molecular insight into the ligand binding site interactions. It appears that the key active site interactions are quite different from the earlier series of inhibitors. Further design of improved reversible SARS-CoV PLpro inhibitors is currently underway in our laboratories.

Experimental Section

Chemistry. ¹H NMR and ¹³C NMR spectra were recorded on Varian Oxford 300 and Bruker Avance 400 spectrometers. Optical rotations were recorded on a Perkin-Elmer 341 polarimeter. Anhydrous solvent was obtained as follows: CH_2Cl_2 by distillation from CaH_2 , THF by distillation from Na and benzophenone. All other solvents were reagent grade. Column chromatography was performed with Whatman 240–400 mesh silica gel under a low pressure of 3–5 psi. TLC was carried out with E. Merck silica gel 60-F-254 plates. Purity of all test compounds was determined by HRMS and HPLC analysis in the different solvent systems. All test compounds showed ≥95% purity.

1-(tert-Butoxycarbonyl)-4-[(3-methoxybenzylamino)carbonyl]piperidine (6b). To a solution of 1-(tert-butoxycarbonyl)piperidine-4-carboxylic acid (344 mg, 1.5 mmol) in dry CH2Cl2 (5 mL), 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride (EDC·HCl) (287 mg, 1.5 mmol), 1-hydroxybenzotriazole hydrate (HOBt·H2O) (203 mg, 1.5 mmol), N-methylmorpholine (NMM) (0.16 mL, 1.5 mmol), and 3-methoxybenzylamine (0.13 mL, 1 mmol) were added successively at 23 °C under argon atmosphere, and the resulting reaction mixture was stirred for 5 h at the same temperature. The reaction mixture was quenched with aqueous NaOH solution and extracted with CH₂Cl₂. The organic layers were dried over anhydrous Na₂SO₄, filtered, and concentrated under reduced pressure. The residue was purified by silica gel column chromatography (40% EtOAc/ hexanes) to furnish **6b** (327 mg, 94%) as a viscous liquid. ¹H NMR (400 MHz, CDCl₃): δ 7.24 (t, J = 7.6 Hz, 1H), 6.76–6.86 (m, 3H), 5.78 (br, 1H), 4.41 (d, J = 5.6 Hz, 2H), 4.12 (br, 2H), 3.79 (s, 3H), 2.73 (br t, J = 11.2 Hz, 2H), 2.25 (tt, J = 4.0 and 11.6 Hz, 1H), 1.82 (br d, J = 12.0 Hz, 2H), 1.65 (ddd, J = 4.1, 12.2, and 24.8 Hz, 2H), 1.45 (s, 9H). ¹³C NMR (100 MHz, CDCl₃): 8 174.1, 159.9, 154.6, 139.7, 129.8, 119.9, 113.4, 112.9, 79.6, 55.2, 43.5, 43.4, 28.6, 28.4.

1-(*tert*-**Butoxycarbonyl**)-**4-**[(**2-methoxybenzylamino**)**carbonyl**]**piperidine** (**6a**). The title compound **6a** was obtained as described for compound 1-(*tert*-butoxycarbonyl)-4-[(3-methoxybenzylamino)carbonyl]piperidine in 92% yield (viscous liquid). ¹H NMR (400 MHz, CDCl₃): δ 7.22 (br t, J = 7.2 Hz, 2H), 6.83-6.92 (m, 2H), 6.09 (br, 1H), 4.41 (d, J = 5.8 Hz, 2H), 4.09 (br, 2H), 3.83 (s, 3H), 2.70 (br t, J = 11.1 Hz, 2 H), 2.20 (tt, J = 3.7 and 11.6 Hz, 1H), 1.77 (br d, J = 12.0 Hz, 2H), 1.59 (ddd, J = 4.4, 12.0, and 24.8 Hz, 2H), 1.43 (s, 9H). ¹³C NMR (100 MHz, CDCl₃): δ 173.9, 157.5, 154.6, 129.6, 128.8, 126.1, 120.6, 110.3, 79.5, 55.3, 43.2, 39.2, 28.5, 28.3.

1-[(1-Naphthyl)methyl]-4-[(3-methoxybenzylamino)carbonyl]piperidine (7b). To a solution of 1-(*tert*-butoxycarbonyl)-4-[(3-methoxybenzylamino)carbonyl]piperidine (100 mg, 0.287 mmol) in CH₂Cl₂ (3 mL), trifluoroacetic acid (0.15 mL) was added at 0 °C. The resulting mixture was stirred for 6 h at 23 °C. The reaction mixture was diluted with CH₂Cl₂ and basified by slow addition of saturated NaHCO3 solution. The layers were separated and the aqueous layer was extracted several times with ethyl acetate. The combined organic layers were dried over anhydrous Na₂SO₄. The solvent was removed under reduced pressure to furnish the amine. To the crude amine in dry CH2Cl2 (5 mL), 1-naphthaldehyde (77 µL, 0.57 mmol), Na(OAc)₃BH (121 mg, 0.57 mmol), and AcOH (33 µL, 0.57 mmol) were added successively at 23 °C, and the resulting mixture was stirred for 12 h at 23 °C. The reaction mixture was basified with 2 N NaOH and diluted with CH₂Cl₂ and H₂O. The organic layer was separated and the aqueous layer extracted with CH₂Cl₂. The combined organic layers were dried over anhydrous Na₂SO₄. Solvent was removed under reduced pressure and the resulting residue was purified by column chromatography over silica gel (2% MeOH/CH₂Cl₂) to provide 1-[(1-naphthyl)methyl]-4-[(3methoxybenzylamino)carbonyl]piperidine as a viscous liquid (79 mg, 71%). ¹H NMR (400 MHz, CDCl₃): δ 8.28–8.33 (m, 1H), 7.82-7.88 (m, 1H), 7.77 (dd, J = 2.2 and 7.1 Hz, 1H), 7.44-7.53 (m, 2H), 7.36-7.43 (m, 2H), 7.23 (t, J = 7.8 Hz, 1H),6.77-6.86 (m, 3H), 5.79 (br, 1H), 4.40 (d, J = 5.7 Hz, 2H), 3.88(s, 2H), 3.78 (s, 3H), 2.94-3.04 (m, 2H), 2.15 (tt, J = 4.2 and 11.4 Hz, 1 H), 2.06 (dt, J = 2.7 and 11.3 Hz, 2H), 1.72–1.88 (m, 4H). ¹³C NMR (100 MHz, CDCl₃): δ 174.9, 159.8, 139.9, 134.3, 133.8, 132.5, 129.7, 128.3, 127.8, 127.2, 125.7, 125.6, 125.0, 124.8, 119.9, 113.3, 112.9, 61.3, 55.2, 53.3, 43.6, 43.3, 29.1. IR (neat): 3290, 2922, 1644, 1598,1263 cm⁻¹. MS (ESI): m/z 389 $[M + H]^+$.

1-[(1-Naphthyl)methyl]-4-[(2-methoxybenzylamino)carbonyl]piperidine (7a). The title compound **7a** was obtained as described for compound **7b** in 70% yield (viscous liquid). ¹H NMR (400 MHz, CDCl₃): δ 8.30 (d, J = 7.9 Hz, 1H), 7.84 (d, J = 7.1 Hz, 1H), 7.77 (d, J = 7.1 Hz, 1H), 7.44–7.53 (m, 2H), 7.37–7.43 (m, 2H), 7.21–7.30 (m, 2H), 6.83–6.94 (m, 2H), 5.98 (br s, 1H), 4.43 (d, J = 5.6 Hz, 2H), 3.87 (s, 2H), 3.84 (s, 3H), 2.98 (d, J = 11.2Hz, 2H), 2.01–2.20 (m, 3H), 1.68–1.84 (m, 4H). ¹³C NMR (100 MHz, CDCl₃): δ 174.6, 157.5, 134.3, 133.8, 132.5, 129.8, 128.8, 128.3, 127.8, 127.2, 126.3, 125.7, 125.6, 125.1, 124.8, 120.7, 110.3, 61.3, 55.3, 53.4, 43.6, 39.3, 29.0. IR (neat): 3305, 1643, 1600, 1242 cm⁻¹. MS (ESI): m/z 389 [M + H]⁺.

1-[(R)-1-(1-Naphthyl)ethyl]-4,4-bis(methoxycarbonyl)-1,4dihydropyridine (12a). A solution of malonate 10 (1.8 g, 5.92 mmol) in 10% hydrochloric acid solution (35 mL) and THF (35 mL) was stirred for 18 h at 23 °C. The solution was neutralized with powdered sodium hydrogen carbonate, and then 1-(R)naphthylmethylamine 11a (1.0 g, 5.84 mmol) in THF (5 mL) was added. After the mixture was stirred for 16 h at 23 °C, the aqueous layer was extracted with EtOAc and dried over Na₂SO₄. Removal of the solvent afforded the residue, which was purified by silica gel column chromatography to furnish compound **12a** (1.1 g, 54%) as a colorless oil. $R_f = 0.74$ (hexane/ EtOAc = 1:1). $[\alpha]^{20}_{D} - 58$ (c 1, CHCl₃). ¹H NMR (300 MHz, CDCl₃): δ 7.90 (d, 1H, J = 7.8 Hz), 7.84 (d, 1H, J = 7.8 Hz), 7.80-7.75 (m, 1H), 7.54-7.40 (m, 4H), 6.21 (d, 2H, J = 8.3 Hz),5.16 (q, 1H, J = 6.6 Hz), 4.77 (d, 2H, J = 8.3 Hz), 3.69 (s, 6H),1.67 (d, 3H, J = 6.6 Hz). ¹³C NMR (75 MHz, CDCl₃): δ 171.4, 136.2, 133.7, 130.8, 129.2, 128.7, 128.4, 126.3, 125.5, 124.9, 123.7, 122.8, 95.3, 56.8, 54.0, 52.4, 19.4. IR (neat): 2951, 1736, 1249, 1069 cm⁻¹. MS (EI): m/z 352 [M + H]⁺. HRMS (EI), calcd for C₂₁H₂₂NO₄ 352.1549, found 352.1553.

1-[(*R*)-**1-**(**2-**Naphthyl)ethyl]-**4**,**4**-bis(methoxycarbonyl)-**1**,**4**dihydropyridine (12b). The title compound was obtained as described in compound **12a** in 58% yield (colorless oil). *R_f* = 0.79 (hexane/EtOAc = 1:1). [α]²⁰_D +32 (*c* 1, CHCl₃). ¹H NMR (300 MHz, CDCl₃): δ 7.84–7.78 (m, 3H), 7.66 (s, 1H), 7.49–7.43 (m, 2H), 7.33 (dd, 1H, *J* = 1.5 and 8.7 Hz), 6.21 (d, 2H, *J* = 8.3 Hz), 4.78 (d, 2H, *J* = 8.3 Hz), 4.59 (q, 1H, *J* = 6.9 Hz), 3.72 (s, 6H), 1.64 (d, 3H, *J* = 6.9 Hz). ¹³C NMR (75 MHz, CDCl₃): δ 171.6, 139.2, 133.1, 132.6, 129.6, 128.4, 127.9, 127.7, 127.5, 126.2, 125.9, 124.8, 95.3, 60.4, 54.1, 52.6, 19.5. IR (neat): 2952, 1732, 1253, 1069 cm⁻¹. MS (EI): *m/z* 292 [M - CO₂Me]⁺. HRMS (EI), calcd for C₁₉H₁₈NO₂ 292.1337, found [M - CO₂Me]⁺ 292.1345.

1-[(*S*)-**1-**(2-Naphthyl)ethyl]-**4**,**4**-bis(methoxycarbonyl)-**1**,**4**dihydropyridine (12c). The title compound was obtained as described in compound **12a** in 54% yield (colorless oil). $R_f =$ 0.73 (hexane/EtOAc = 1:1). [α]²⁰_D -32 (*c* 1, CHCl₃). MS (EI): *m*/*z* 351 [M]⁺. HRMS (EI), calcd for C₂₁H₂₁NO₄ 351.1471, found [M]⁺ 351.1477.

1-[(*S*)-**1-**(**1-Naphthyl**)**ethyl**]-**4**,**4**-**bis**(**methoxycarbonyl**)-**1**,**4**-**dihydropyridine** (**12d**). The title compound was obtained as described in compound **12a** in 42% yield (colorless oil). $R_f = 0.77$ (hexane/EtOAc = 1:1). [α]²⁰_D+57 (*c* 1, CHCl₃). MS (ESI): m/z 374 [M + Na]⁺. HRMS (ESI), calcd for C₂₁H₂₁NO₄Na 374.1368, found 374.1371.

1-(1-Naphthylmethyl)-4,4-bis(methoxycarbonyl)-1,4-dihydropyridine (12e). The title compound was obtained as described in compound **12a** in 39% yield (colorless oil). $R_f = 0.82$ (hexane/ EtOAc = 1:1). ¹H NMR (300 MHz, CDCl₃): δ 7.86–7.80 (m, 2H), 7.77 (d, 1H, J = 8.7 Hz), 7.54–7.48 (m, 2H), 7.42 (t, 1H, J = 8.3 Hz), 7.30 (d, 1H, J = 6.9 Hz), 6.15 (d, 2H, J = 8.3 Hz), 4.82 (d, 2H, J = 8.3 Hz), 4.74 (s, 2H), 3.73 (s, 6H). ¹³C NMR (75 MHz, CDCl₃): δ 171.6, 133.5, 132.6, 131.1, 130.7, 128.7, 128.2, 126.4, 125.8, 125.4, 125.1, 122.5, 95.3, 54.5, 53.7, 52.7. IR (neat): 2951, 1735, 1253, 1067 cm⁻¹. MS (EI): m/z 278 [M – CO₂Me]⁺. HRMS (EI), calcd for C₁₈H₁₆NO₂ 278.1181, found 278.1185.

1-(2-Naphthylmethyl)-4,4-bis(methoxycarbonyl)-1,4-dihydropyridine (12f). The title compound was obtained as described in compound **12a** in 62% yield (colorless oil). $R_f = 0.80$ (hexane/ EtOAc = 1:1). ¹H NMR (300 MHz, CDCl₃): δ 7.80–7.77 (m, 3H), 7.60 (s, 1H), 7.48–7.41 (m, 2H), 7.28 (d, 1H, J = 1.8 Hz), 6.16 (d, 2H, J = 8.0 Hz), 4.81 (d, 2H, J = 8.0 Hz), 4.41 (s, 2H), 3.73 (s, 6H). ¹³C NMR (75 MHz, CDCl₃): δ 171.5, 134.9, 133.1, 132.6, 131.2, 128.5, 127.7, 127.5, 126.2, 125.9, 125.8, 124.8, 95.3, 56.9, 53.6, 52.6. IR (neat): 2950, 1731, 1253, 1066 cm⁻¹. MS (EI): m/z 278 [M - CO₂Me]⁺. HRMS (EI), calcd for C₁₈H₁₆NO₂ 278.1181, found 278.1184.

1-[1-Methyl-1-(1-naphthyl)ethyl]-4,4-bis(methoxycarbonyl)-1,4-dihydropyridine (12g). The title compound was obtained as described in compound **12a** in 41% yield (colorless oil). $R_f = 0.77$ (hexane/EtOAc = 1:1). ¹H NMR (300 MHz, CDCl₃): δ 8.20–8.16 (m, 1H), 7.89 (d, 1H, J = 7.8 Hz), 7.84–7.80 (m, 1H), 7.77 (d, 1H, J = 7.8 Hz), 7.52–7.36 (m, 4H), 6.27 (d, 2H, J = 8.1 Hz), 4.77 (d, 2H, J = 8.1 Hz), 3.69 (s, 6H), 1.77 (s, 6H). ¹³C NMR (75 MHz, CDCl₃): δ 171.6, 140.1, 134.7, 130.5, 129.1, 129.0, 127.7, 126.1, 126.0, 125.3, 124.7, 124.0, 96.0, 61.9, 53.8, 52.5, 28.7. IR (neat): 2951, 1736, 1252, 1062 cm⁻¹. MS (ESI): m/z 388 [M + Na]⁺. HRMS (ESI), calcd for C₂₂H₂₃NO₄Na 388.1525, found 388.1529.

1-[(R)-1-(1-Naphthyl)ethyl]-4,4-bis(methoxycarbonyl)piperidine (13a). To a stirred solution of dihydropyridine 12a (1.1 g, 3.1 mmol) in EtOAc (75 mL) was added platinum(IV) oxide (100 mg), and the mixture was allowed to stir for 2 h at 23 °C under H₂ atmosphere. The mixture was filtered through Celite pad, and the filtrate was concentrated under reduced pressure. The residue was purified by silica gel column chromatography to furnish compound 13a (942 mg, 88%) as a colorless oil. $R_f = 0.7$ (hexane/EtOAc = 1:1). $[\alpha]_{D}^{20}$ +9 (*c* 1, CHCl₃). ¹H NMR (300 MHz, CDCl₃): δ 8.44-8.41 (m, 1H), 7.85-7.81 (m, 1H), 7.73 (d, 1H, J = 8.1 Hz), 7.56 (d, 1H, J = 6.9 Hz), 7.50–7.39 (m, 3H), 4.03 7.73 (q, 1H, J = 6.3 Hz), 3.72 (s, 6H), 2.58-2.56 (m, 2H), 2.47-2.40 (m, 2H), 2.20-2.04 (m, 4H), 1.44 (d, 3H, J = 6.3 Hz). ¹³C NMR (75 MHz, CDCl₃): δ 171.8, 140.7, 134.0, 131.5, 128.6, 127.3, 125.4, 125.3, 125.2, 124.5, 124.1, 61.7, 53.3, 52.4, 47.9, 31.2, 18.7. IR (neat): 2952, $1734, 1251 \text{ cm}^{-1}$. MS (ESI): $m/z 356 [M + H]^+$. HRMS (ESI), calcd for C₂₁H₂₆NO₄ 356.1862, found 356.1866.

1-[(R)-1-(2-Naphthyl)ethyl]-4,4-bis(methoxycarbonyl)piperidine (13b). The title compound was obtained as described in compound **13a** in 74% yield (colorless oil). $R_f = 0.48$ (hexane/ EtOAc = 1:1). [α]²⁰_D +23 (*c* 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 7.83–7.80 (m, 3H), 7.72 (s, 1H), 7.53 (dd, 1H, *J* = 1.3 and 8.7 Hz), 7.49–7.42 (m, 2H), 3.73 (s, 6H), 3.49 (q, 1H, *J* = 6.7 Hz), 2.57–2.55 (bm, 2H), 2.47–2.42 (m, 2H), 2.24–2.13 (m, 4H), 1.42 (d, 3H, *J* = 6.7 Hz). ¹³C NMR (100 MHz, CDCl₃): δ 171.7, 142.1, 133.3, 132.7, 128.0, 127.7, 127.5, 125.8, 125.4, 64.8, 53.2, 52.5, 47.9, 31.1, 19.6. IR (neat): 2951, 1731, 1250, 1073 cm⁻¹. MS (EI): *m*/*z* 355 [M]⁺. HRMS (EI), calcd for C₂₁H₂₅NO₄ 355.1784, found 355.1781.

1-[(*S*)-**1-**(2-Naphthyl)ethyl]-4,4-bis(methoxycarbonyl)piperidine (13c). The title compound was obtained as described in compound 13a in 67% yield (colorless oil). $R_f = 0.48$ (hexane/EtOAc = 1:1). [α]²⁰_D -24 (*c* 1, CHCl₃). MS (EI): *m/z* 355 [M]⁺. HRMS (EI), calcd for C₂₁H₂₅NO₄ 355.1784, found 355.1786.

1-[(*S*)-**1-**(**1-Naphthyl**)**ethyl**]-**4**,**4**-**bis**(**methoxycarbonyl**)**piperidine** (**13d**). The title compound was obtained as described in compound **13a** in 87% yield (colorless oil). $R_f = 0.7$ (hexane/EtOAc = 1:1). [α]²⁰_D -9 (c 1, CHCl₃). MS (ESI): m/z 356 [M + H]⁺. HRMS (ESI), calcd for C₂₁H₂₆NO₄ 356.1862, found 356.1865.

1-(1-Naphthylmethyl)-4,4-bis(methoxycarbonyl)piperidine (13e). The title compound was obtained as described in compound **13a** in 60% yield (colorless oil). $R_f = 0.70$ (hexane/EtOAc = 1:1). ¹H NMR (300 MHz, CDCl₃): δ 8.29–8.26 (m, 1H), 7.84–7.81 (m, 1H), 7.77–7.73 (m, 1H), 7.52–7.43 (m, 2H), 7.38–7.36 (m, 2H), 3.84 (s, 2H), 3.72 (s, 6H), 2.48 (t, 4H, J = 5.4 Hz), 2.13 (t, 4H, J = 5.4 Hz). ¹³C NMR (75 MHz, CDCl₃): δ 171.7, 134.1, 133.8, 132.5, 128.3, 127.9, 127.2, 125.6, 125.5, 125.0, 124.8, 61.2, 53.3, 52.5, 50.6, 31.0. IR (neat): 2950, 1732, 1254, 1072 cm⁻¹. MS (EI): m/z 341 [M]⁺. HRMS (EI), calcd for C₂₀H₂₃NO₄ 341.1627, found 341.1630.

1-(2-Naphthylmethyl)-4,4-bis(methoxycarbonyl)piperidine (13f). The title compound was obtained as described in compound **13a** in 94% yield (colorless oil). $R_f = 0.48$ (hexane/EtOAc = 1:1). ¹H NMR (400 MHz, CDCl₃): δ 7.83–7.78 (m, 3H), 7.72 (s, 1H), 7.50–7.42 (m, 3H), 3.74 (s, 6H), 3.61 (s, 2H), 2.48 (bm, 4H), 2.19 (t, 4H, J = 5.5 Hz). ¹³C NMR (100 MHz, CDCl₃): δ 171.7, 135.9, 133.2, 132.7, 127.8, 127.6, 127.6, 127.5, 127.2, 125.9, 125.5, 63.2, 53.2, 52.5, 50.5, 30.9. IR (neat): 2950, 1732, 1254, 1073 cm⁻¹. MS (EI): m/z 341 [M]⁺. HRMS (EI), calcd for C₂₀H₂₃NO₄ 341.1627, found [M]⁺ 341.1626.

1-[1-Methyl-1-(1-naphthyl)ethyl]-4,4-bis(methoxycarbonyl)piperidine (13g). The title compound **13g** was obtained as described in compound **13a** in 93% yield (colorless oil). $R_f = 0.29$ (hexane/ EtOAc = 9:1). ¹H NMR (400 MHz, CDCl₃): δ 9.59 (d, 1H, J = 7.9 Hz), 7.84 (dd, 1H, J = 2.2 and 7.0 Hz), 7.74 (d, 1H, J = 8.0 Hz), 7.52–7.44 (m, 3H), 7.37 (t, 1H, J = 7.7 Hz), 3.74 (s, 6H), 2.63 (bm, 4H), 2.14 (bm, 4H), 1.59 (s, 6H). ¹³C NMR (100 MHz, CDCl₃): δ 171.8, 143.7, 134.9, 132.0, 128.6, 128.2, 128.0, 125.1, 124.7, 124.4, 123.6, 62.2, 53.6, 52.4, 43.4, 31.8, 23.6. IR (neat): 2973, 1735, 1251, 1124, 780 cm⁻¹. MS (ESI): m/z 370 [M + H]⁺. HRMS (ESI), calcd for C₂₂H₂₈NO₄ 370.2018, found 370.2013.

1-[(R)-1-(1-Naphthyl)ethyl]-4-methoxycarbonylpiperidine (14a). To a stirred solution of dimethyl malonate 13a (917 mg, 2.58 mmol) in DMF (25 mL) was added sodium cyanide (190 mg, 3.88 mmol) at 23 °C, and the mixture was allowed to stir for 16 h at reflux temperature. The mixture was diluted with EtOAc and washed with water. The organic layer was dried over Na₂SO₄, filtered, and concentrated. The residue was purified by silica gel column chromatography to furnish compound 14a (704 mg, 92%) as a colorless oil. $R_f = 0.56 (CH_2Cl_2/MeOH = 9:1). [\alpha]^{20}_{D} + 2$ (c 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 8.48 (dd, 1H, J = 1.2 and 7.6 Hz), 7.87 (d, 1H, J = 7.1 Hz), 7.76 (d, 1H, J = 8.1 Hz), 7.60 (d, 1H, J = 7.1 Hz), 7.53 - 7.43 (m, 3H), 4.12 (q, 1H, J = 6.7)Hz), 3.68 (s, 3H), 3.17-3.15 (m, 1H), 2.87-2.84 (m, 1H), 2.35-2.27 (m, 1H), 2.10 (ddd, 2H, J = 2.6, 11.2, and 19.8 Hz), 1.97-1.92 (m, 1H), 1.83-1.71 (m, 3H), 1.49 (d, 3H, J = 6.7 Hz).¹³C NMR (100 MHz, CDCl₃): δ 175.8, 140.8, 134.0, 131.6, 128.6, 127.2, 125.4, 125.3, 125.3, 124.5, 124.2, 61.6, 51.5, 49.1, 41.3, 28.7, 28.6, 18.6. IR (neat): 2950, 1732, 1169, 780 cm⁻¹. MS (EI): m/z 297 [M]⁺. HRMS (EI), calcd for C₁₉H₂₃NO₂ 297.1729, found 297.1730.

1-[(*R*)-**1-**(2-Naphthyl)ethyl]-4-methoxycarbonylpiperidine (14b). The title compound was obtained as described in compound **14a** in 78% yield (colorless oil). $R_f = 0.43$ (CH₂Cl₂/MeOH = 9:1). [α]²⁰_D +16 (*c* 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 7.84–7.80 (m, 3H), 7.72 (s, 1H), 7.53 (dd, 1H, *J* = 1.1 and 8.4 Hz), 7.50–7.43 (m, 2H), 3.67 (s, 3H), 3.57 (q, 1H, *J* = 6.8 Hz), 3.08–3.06 (m, 1H), 2.86–2.83 (m, 1H), 2.29–2.22 (m, 1H), 2.09–1.91 (m, 3H), 1.87–1.71 (m, 3H), 1.45 (d, 3H, *J* = 6.8 Hz). ¹³C NMR (100 MHz, CDCl₃): δ 175.7, 141.7, 133.3, 132.7, 127.8, 127.7, 127.5, 126.0, 125.9, 125.8, 125.4, 64.7, 51.5, 50.5, 49.6, 41.2, 28.5, 19.3. IR (neat): 2949, 1732, 1258, 1172 cm⁻¹. MS (EI): *m/z* 297 [M]⁺. HRMS (EI), calcd for C₁₉H₂₃NO₂ 297.1729, found [M]⁺ 297.1732.

1-[(*S*)-1-(2-Naphthyl)ethyl]-4-methoxycarbonylpiperidine (14c). The title compound was obtained as described in compound 14a in 90% yield (colorless oil). $R_f = 0.47$ (CH₂Cl₂/MeOH = 9:1). $[\alpha]^{20}_{D} - 15$ (*c* 1, CHCl₃). MS (EI): m/z 297 [M]⁺. HRMS (EI), calcd for C₁₉H₂₃NO₂ 297.1729, found 297.1731.

1-[(*S*)-1-(1-Naphthyl)ethyl]-4-methoxycarbonylpiperidine (14d). The title compound was obtained as described in compound 14a in 76% yield (colorless oil). $R_f = 0.57$ (CH₂Cl₂/MeOH = 9:1). $[\alpha]^{20}_{D} - 2 (c 1, \text{CHCl}_3)$. MS (EI): *m*/*z* 297 [M]⁺. HRMS (EI), calcd for C₁₉H₂₃NO₂ 297.1729, found 297.1729.

1-(1-Naphthylmethyl)-4-methoxycarbonylpiperidine (14e). The title compound was obtained as described in compound **14a** in 38% yield (colorless oil). $R_f = 0.53$ (CH₂Cl₂/MeOH = 9:1). ¹H NMR (400 MHz, CDCl₃): δ 8.31 (d, 1H, J = 7.8 Hz), 7.86 (dd, 1H, J = 1.6 and 7.1 Hz), 7.78 (d, 1H, J = 7.1 Hz), 7.53–7.47 (m, 2H), 7.44–7.39 (m, 2H), 3.89 (s, 2H), 3.68 (s, 3H), 2.94–2.88 (m, 2H), 2.37–2.30 (m, 1H), 2.12 (dt, 2H, J = 1.6 and 11.2 Hz), 1.90–1.86 (m, 2H), 1.81–1.72 (m, 2H). ¹³C NMR (100 MHz, CDCl₃): δ 175.7, 134.2, 133.7, 132.5, 128.3, 127.8, 127.2, 125.6, 125.5, 125.0, 124.7, 61.3, 53.1, 51.5, 41.1, 28.3. IR (neat): 2949, 1736, 1167, 788 cm⁻¹. MS (EI): m/z 283 [M]⁺. HRMS (EI), calcd for C₁₈H₂₁NO₂ 283.1572, found 283.1569.

1-(2-Naphthylmethyl)-4-methoxycarbonylpiperidine (14f). The title compound was obtained as described in compound **14a** in 47% yield (colorless oil). $R_f = 0.44$ (CH₂Cl₂/MeOH = 9:1). ¹H NMR (400 MHz, CDCl₃): δ 7.84–7.80 (m, 3H), 7.73 (s, 1H), 7.51–7.43 (m, 3H), 3.68 (s, 3H), 3.65 (s, 2H), 2.93–2.88 (m, 2H), 2.36–2.28 (m, 1H), 2.08 (dt, 2H, J = 2.2 and 11.4 Hz), 1.94–1.75 (m, 4H). ¹³C NMR (100 MHz, CDCl₃): δ 175.6, 135.9, 133.2, 132.7, 127.8, 127.6, 127.6, 127.5, 127.3, 125.9, 125.5, 63.3, 52.9, 51.6, 41.0, 28.2. IR (neat): 2948, 1736, 1167 cm⁻¹. MS (ESI): m/z 284 [M + H]⁺. HRMS (ESI), calcd for C₁₈H₂₂NO₂ 284.1651, found 284.1652.

1-[1-Methyl-1-(1-naphthyl)ethyl]-4-methoxycarbonylpiperidine (14g). The title compound was obtained as described in compound 14a in 87% yield (colorless oil). $R_f = 0.43$ (hexane/ EtOAc = 9:1). ¹H NMR (400 MHz, CDCl₃): δ 9.63–9.60 (m, 1H), 7.87–7.84 (m, 1H), 7.76 (d, 1H, J = 8.0 Hz), 7.51–7.46 (m, 3H), 7.39 (t, 1H, J = 7.8 Hz), 3.69 (s, 3H), 2.96 (bs, 2H), 2.34–2.28 (m, 3H), 1.84–1.75 (bm, 4H), 1.61 (s, 6H). ¹³C NMR (100 MHz, CDCl₃): δ 175.9, 144.1, 134.8, 132.0, 128.6, 128.4, 127.9, 125.1, 124.7, 124.4, 123.5, 62.3, 51.5, 45.9, 41.9, 29.2, 22.7. IR (neat): 2950, 1736, 1171, 780 cm⁻¹. MS (EI): m/z 311 [M]⁺. HRMS (EI), calcd for C₂₀H₂₅NO₂ 311.1885, found 311.1891.

1-[(*R*)-1-(1-Naphthyl)ethyl]-4-(2-methoxybenzylamino)carbonylpiperidine (15a). To a stirred solution of ester 14a (106 mg, 0.36 mmol) in THF/MeOH/H₂O (3:1:1) (8 mL) was added LiOH \cdot H₂O (22.4 mg, 0.53 mmol) at 0 °C, and the mixture was allowed to stir for 16 h at 23 °C. The mixture was concentrated, and to it was added a saturated NaHCO₃ solution. The mixture was extracted with Et₂O. The aqueous layer was adjusted to pH 2 with 10% HCl solution and extracted with EtOAc. The organic layers were dried over Na₂SO₄, filtered, and concentrated to give the corresponding acid as a colorless oil. To a solution of acid (0.36 mmol), N-(3-dimethylaminopropyl)-N'-ethylcarbodiimide hydrochloride (EDCI) (138.0 mg, 0.72 mmol) and 1-hydroxybenzotriazole hydrate (HOBT) (97.3 mg, 0.72 mmol) in dry CH₂Cl₂/DMF (9:1) (8 mL) were added 2-methoxybenzylamine 5a (52.7 μ L, 0.40 mmol) and diisopropylethylamine (0.38 mL, 2.2 mmol) at 0 °C under argon atmosphere. The mixture was allowed to stir for 15 h at 23 °C. The reaction mixture was quenched with water and extracted with CH₂Cl₂. The organic layers were dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The residue was purified by silica gel column chromatography to furnish compound 15a (143 mg, 99%) as a white amorphous solid. $R_f = 0.42$ (CH₂Cl₂/MeOH = 9:1). [α]²⁰_D -2 (*c* 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 8.46 (d, 1H, J = 7.8 Hz), 7.86-7.84 (m, 1H), 7.74 (d, 1H, J = 8.1 Hz), 7.58 (d, 1H, J =7.0 Hz), 7.51-7.41 (m, 3H), 7.23 (d, 1H, J = 7.3 Hz), 6.90 (t, 1H, J = 7.3 Hz), 6.85 (d, 1H, J = 8.4 Hz), 6.17 (bt, 1H, J = 5.8 Hz), 4.44 (d, 2H, J = 5.8 Hz), 4.10 (q, 1H, J = 6.6 Hz), 3.81 (s, 3H),3.23-3.20 (m, 1H), 2.87-2.85 (m, 1H), 2.12-1.95 (m, 3H), 1.89-1.68 (m, 4H), 1.47 (d, 3H, J = 6.6 Hz). ¹³C NMR (100 MHz, CDCl₃): δ 174.9, 157.5, 140.9, 134.1, 131.7, 129.6, 128.7, 128.7, 127.3, 126.5, 125.5, 125.4, 125.3, 124.5, 124.3, 120.7, 110.3, 61.7, 55.3, 52.0, 49.2, 43.7, 39.1, 29.3, 18.7. IR (neat): 3302, 2938, $1650, 1243, 753 \text{ cm}^{-1}$. MS (ESI): $m/z 403 [M + H]^+$. HRMS (ESI), calcd for C₂₆H₃₁N₂O₂ 403.2386, found 403.2388.

1-[(*R*)-**1-**(**1-Naphthyl**)**ethyl**]-**4-**(**3-methoxybenzylamino**)**carbo-nylpiperidine** (**15b**). The title compound was obtained as described in compound **15a** in 95% yield (white amorphous solid). $R_f = 0.49$ (CH₂Cl₂/MeOH = 9:1). [α]²⁰_D - 2 (*c* 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 8.45 (d, 1H, *J* = 7.7 Hz), 7.86–7.84 (m, 1H), 7.74 (d, 1H, *J* = 8.1 Hz), 7.58 (d, 1H, *J* = 7.1 Hz), 7.51–7.41 (m, 3H), 6.83–6.78 (m, 3H), 6.12 (bt, 1H, *J* = 5.7 Hz), 4.37 (d, 2H, *J* = 5.7 Hz), 4.10 (q, 1H, *J* = 6.6 Hz), 3.76 (s, 3H), 3.23–3.20 (m, 1H), 2.90–2.85 (m, 1H), 2.14–1.95 (m, 3H), 1.89–1.69 (m, 4H), 1.47 (d, 3H, *J* = 6.6 Hz). ¹³C NMR (100 MHz, CDCl₃): δ 175.2, 159.8, 140.7, 140.1, 134.0, 131.6, 129.6, 128.6, 127.2, 125.4, 125.3, 125.3, 124.5, 124.2, 119.8, 113.2, 112.7, 61.6, 55.1, 51.8, 49.1, 43.5, 43.2, 29.3, 18.6. IR (neat): 3293, 2937, 1644, 1263, 781 cm⁻¹. MS (EI): *m/z* 402 [M]⁺. HRMS (EI), calcd for C₂₆H₃₀N₂O₂ 402.2307, found 402.2303.

1-[(R)-1-(1-Naphthyl)ethyl]-4-(4-methoxybenzylamino)carbonylpiperidine (15c). The title compound was obtained as described in compound 15a in 88% yield (white amorphous solid). $R_f = 0.56$ $(CH_2Cl_2/MeOH = 9:1). [\alpha]_{D}^{20} - 2 (c 1, CHCl_3).$ ¹H NMR (400) MHz, CDCl₃): δ 8.45 (d, 1H, J = 7.7 Hz), 7.86–7.84 (m, 1H), 7.74 (d, 1H, J = 8.1 Hz), 7.57 (d, 1H, J = 7.1 Hz), 7.51-7.41 (m, J = 7.1 Hz), 7.51-7.51 (m, J = 7.51 (m, J = 7.51), 7.51-7.51 (m, J = 7.51), 7.51 (m, J = 73H), 7.27 (d, 2H, J = 8.5 Hz), 6.84 (d, 2H, J = 8.5 Hz), 5.97 (bt, 1H, J = 5.6 Hz), 4.33 (d, 2H, J = 5.6 Hz), 4.10 (q, 1H, J = 6.7Hz), 3.77 (s, 3H), 3.23-3.20 (m, 1H), 2.89-2.86 (m, 1H), 2.13-1.95 (m, 3H), 1.89-1.68 (m, 4H), 1.47 (d, 3H, J = 6.7Hz). ¹³C NMR (100 MHz, CDCl₃): δ 175.0, 158.9, 140.7, 134.0, 131.6, 130.5, 129.0, 128.6, 127.2, 125.4, 125.3, 125.3, 124.5, 124.2, 114.0, 61.6, 55.2, 51.8, 49.1, 43.5, 42.8, 29.3, 18.6. IR (neat): 3292, 2932, 1513, 1644, 1249, 781 cm⁻¹. MS (EI): *m/z* 402 [M]⁺. HRMS (EI), calcd for C₂₆H₃₀N₂O₂ 402.2307, found 402.2299.

1-[(*R*)-**1-**(2-Naphthyl)ethyl]-**4-**(3-methoxybenzylamino)carbonylpiperidine (15d). The title compound was obtained as described in compound **15a** in 94% yield (white amorphous solid). $R_f = 0.43$ (CH₂Cl₂/MeOH = 9:1). [α]²⁰_D +10 (*c* 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 7.82-7.79 (m, 3H), 7.70 (s, 1H), 7.52 (dd, 1H, J = 1.2 and 8.4 Hz), 7.49-7.42 (m, 2H), 7.22 (t, 1H, J = 7.6 Hz), 6.83-6.79 (m, 3H), 5.95 (bt, 1H, J = 5.7 Hz), 4.38 (d, 2H, J = 5.7 Hz), 3.77 (s, 3H), 3.56 (q, 1H, J = 6.7 Hz), 3.16-3.14 (m, 1H), 2.90-2.88 (m, 1H), 2.11-1.91 (m, 3H), 1.89-1.70 (m, 4H), 1.44 (d, 3H, J = 6.7 Hz). ¹³C NMR (100 MHz, CDCl₃): δ 175.0, 159.8, 141.7, 140.0, 133.2, 132.6, 129.6, 127.8, 127.7, 127.5, 125.9, 125.9, 125.8, 125.4, 119.8, 113.2, 112.8, 64.7, 55.1, 50.8, 49.7, 43.4, 43.2, 29.1, 19.3. IR (neat): 3296, 2932, 1645, 1264 cm⁻¹. MS (ESI): *m/z* 403 [M + H]⁺. HRMS (ESI), calcd for C₂₆H₃₁N₂O₂ 403.2386, found 403.2390.

1-[(*R*)-**1-**(2-Naphthyl)ethyl]-4-(2-methoxybenzylamino)carbonylpiperidine (15e). The title compound was obtained as described in compound **15a** in 98% yield (white amorphous solid). $R_f = 0.47$ (CH₂Cl₂/MeOH = 9:1). [α]²⁰_D + 12 (*c* 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 7.82–7.79 (m, 3H), 7.70 (s, 1H), 7.51 (d, 1H, J = 8.4 Hz), 7.49–7.43 (m, 2H), 7.28–7.24 (m, 2H), 6.93–6.85 (m, 2H), 6.02 (bt, 1H, J = 5.7 Hz), 4.43 (d, 2H, J = 5.7 Hz), 3.84 (s, 3H), 3.57 (q, 1H, J = 6.7 Hz), 3.16–3.14 (m, 1H), 2.90–2.88 (m, 1H), 2.08–1.86 (m, 4H), 1.84–1.64 (m, 3H), 1.45 (d, 3H, J = 6.7 Hz). ¹³C NMR (100 MHz, CDCl₃): δ 174.6, 157.5, 141.6, 133.2, 132.6, 129.7, 129.0, 128.8, 128.2, 127.8, 127.7, 127.5, 126.3, 126.0, 125.8, 125.4, 120.6, 110.2, 64.7, 55.2, 50.8, 49.8, 43.5, 39.2, 29.1, 19.3. IR (neat): 3306, 2933, 1645, 1243, 751 cm⁻¹. MS (ESI): *m*/*z* 403 [M + H]⁺. HRMS (ESI), calcd for C₂₆H₃₁N₂O₂ 403.2386, found 403.2394.

1-[(*S*)-**1-**(2-Naphthyl)ethyl]-**4-**(3-methoxybenzylamino)carbonylpiperidine (15f). The title compound was obtained as described in compound **15a** in 83% yield (white amorphous solid). $R_f = 0.37 (CH_2Cl_2/MeOH = 9:1). [α]^{20}_D - 10 (c 1, CHCl_3). MS$ (ESI): m/z 403 [M + H]⁺. HRMS (ESI), calcd for C₂₆H₃₁N₂O₂ 403.2386, found 403.2392.

1-[(R)-1-(1-Naphthyl)ethyl]-4-[3,4-(methylenedioxy)benzylamino]carbonylpiperidine (15g). The title compound was obtained as described in compound 15a in 93% yield (white amorphous solid). $R_f = 0.47 (CH_2Cl_2/MeOH = 9:1). [\alpha]_{D}^{20}$ -2 (c 1, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ 8.45 (d, 1H, J = 7.5 Hz), 7.86–7.83 (m, 1H), 7.73 (d, 1H, J = 8.1 Hz), 7.56 (d, 1H, J = 6.9 Hz), 7.51 - 7.40 (m, 3H), 6.73 - 6.66 (m, 3H), 6.20(bt, 1H, J = 5.7 Hz), 5.89 (s, 2H), 4.29 (d, 2H, J = 5.7 Hz), 4.08(q, 1H, J = 6.7 Hz), 3.22-3.19 (m, 1H), 2.88-2.85 (m, 1H),2.13-1.94 (m, 3H), 1.87-1.67 (m, 4H), 1.46 (d, 3H, J = 6.7 Hz).¹³C NMR (100 MHz, CDCl₃): δ 175.2, 147.8, 146.8, 140.7, 134.0, 132.4, 131.6, 128.6, 127.2, 125.4, 125.3, 125.3, 124.5, 124.2, 120.8, 108.2, 100.9, 61.6, 51.8, 49.1, 43.5, 43.0, 29.3, 18.6. IR (neat): 3294, 2924, 1644, 1489, 1252, 1040, 781 cm⁻¹. MS (ESI): m/z 417 [M + H]⁺. HRMS (ESI), calcd for C₂₆H₂₉N₂O₃ 417.2178, found 417.2178.

1-[(*S*)-**1-**(**1-Naphthyl**)ethyl]-**4-**[**3,4-**(methylenedioxy)benzylamino]carbonylpiperidine (15h). The title compound was obtained as described in compound **15a** in 80% yield (white amorphous solid). $R_f = 0.56$ (CH₂Cl₂/MeOH = 9:1). [α]²⁰_D +2 (*c* 1, CHCl₃). MS (EI): *m*/*z* 417 [M + H]⁺. HRMS (EI), calcd for C₂₆H₂₉N₂O₃ 417.2178, found 417.2173.

1-(1-Naphthylmethyl)-4-[3,4-(methylenedioxy)benzylamino]carbonylpiperidine (15i). The title compound was obtained as described in compound **15a** in > 99% yield (white amorphous solid). $R_f = 0.48$ (CH₂Cl₂/MeOH = 9:1). ¹H NMR (400 MHz, CDCl₃): δ 8.30–8.28 (m, 1H), 7.84 (d, 1H, J = 7.2 Hz), 7.77 (d, 1H, J = 7.8 Hz), 7.52–7.45 (m, 2H), 7.41–7.37 (m, 2H), 6.74–6.67 (m, 3H), 5.91 (bm, 1H), 5.91 (s, 2H), 4.30 (d, 2H, J = 5.7 Hz), 3.87 (s, 2H), 2.99–2.96 (m, 2H), 2.35–2.29 (m, 1H), 2.18–2.01 (m, 3H), 1.88–1.72 (m, 3H). ¹³C NMR (100 MHz, CDCl₃): δ 174.9, 147.8, 146.8, 134.1, 133.7, 132.5, 132.2, 128.3, 127.8, 127.2, 126.0, 125.6, 125.5, 125.0, 124.7, 120.9, 108.2, 101.0, 61.2, 53.3, 43.4, 43.1, 28.9. IR (neat): 3307, 2924, 1645, 1490, 1252, 1040 cm⁻¹. MS (ESI): m/z 403 [M + H]⁺. HRMS (ESI), calcd for C₂₅H₂₇N₂O₃ 403.2022, found 403.2025.

1-(2-Naphthylmethyl)-4-[3,4-(methylenedioxy)benzylamino]carbonylpiperidine (15j). The title compound was obtained as described in compound **15a** in 88% yield (white amorphous solid). $R_f = 0.42$ (CH₂Cl₂/MeOH = 9:1). ¹H NMR (400 MHz, CDCl₃): δ 7.82–7.78 (m, 3H), 7.72 (s, 1H), 7.49–7.42 (m, 3H), 6.74–6.68 (m, 3H), 6.01 (t, 1H, J = 5.6 Hz), 5.91 (s, 2H), 4.31 (d, 2H, J = 5.6 Hz), 3.63 (s, 2H), 2.97–2.94 (m, 2H), 2.15–2.07 (m, 1H), 2.04–1.98 (m, 2H), 1.83–1.77 (m, 4H). ¹³C NMR (100 MHz, CDCl₃): δ 174.9, 147.8, 146.8, 135.9, 132.7, 132.3, 127.8, 127.6, 127.6, 127.4, 127.3, 125.9, 125.5, 120.9, 108.2, 101.0, 63.2, 53.1, 43.3, 43.1, 28.9. IR (neat): 3293, 2923, 1643, 1490, 1252, 1040 cm^{-1} . MS (ESI): $m/z 403 [M + H]^+$. HRMS (ESI), calcd for $C_{25}H_{27}N_2O_3 403.2022$, found 403.2025.

1-[1-Methyl-1-(1-naphthyl)ethyl]-4-[3,4-(methylenedioxy)benzylamino]carbonylpiperidine (15k). The title compound was obtained as described in compound **15a** in 90% yield (white amorphous solid). $R_f = 0.51$ (hexane/EtOAc = 1:1). ¹H NMR (400 MHz, CDCl₃): δ 9.56–9.53 (m, 1H), 7.82–7.79 (m, 1H), 7.72 (d, 1H, J =7.9 Hz), 7.46–7.41 (m, 3H), 7.36 (t, 1H, J = 7.6 Hz), 6.73–6.71 (m, 2H), 6.67 (dd, 1H, J = 1.1 and 8.2 Hz), 6.00 (bt, 1H, J = 5.7 Hz), 5.90 (s, 2H), 4.29 (d, 2H, J = 5.7 Hz), 2.92 (bs, 2H), 2.21 (bm, 2H), 2.09–2.01 (m, 1H), 1.72 (bm, 4H), 1.56 (s, 6H). ¹³C NMR (100 MHz, CDCl₃): δ 175.2, 147.8, 146.8, 144.0, 134.8, 132.4, 132.0, 128.5, 128.3, 127.9, 125.1, 124.7, 124.4, 123.5, 120.8, 108.2, 100.9, 62.3, 46.1, 44.0, 43.1, 29.8, 24.7. IR (neat): 3294, 2974, 1642, 1490, 1253, 1041, 780 cm⁻¹. MS (ESI): m/z 311 [M + H]⁺. HRMS (ESI), calcd for C₂₇H₃₁N₂O₃ 431.2335, found 431.2330.

1-[1-(1-Naphthyl)ethyl]-4-tert-butoxycarbonylpiperazine (18). To a solution of N-Boc-piperazine 16 (107 mg, 0.58 mmol) and 1-acetonaphthone 17 (0.10 mL, 0.69 mmol) in MeOH/AcOH (50:1) (4 mL) was added sodium cyanoborohydride (38 mg, 0.58 mmol) at 0 °C. The mixture was allowed to stir for 48 h at 23 °C. The reaction was quenched with saturated NaHCO₃ solution, and the organic layer was extracted with CH₂Cl₂. The organic layer was dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The residue was purified by silica gel column chromatography to furnish compound 18 (46 mg, 24%) as a colorless oil. $R_f = 0.75$ (hexane/EtOAc = 1:1). ¹H NMR (300 MHz, CDCl₃): δ 8.42 (d, 1H, J = 7.2 Hz), 7.86–7.83 (m, 1H), 7.74 (d, 1H, J = 10.8 Hz), 7.57 (d, 1H, J = 7.2 Hz), 7.50–7.39 (m, 3H), 4.09 (q, 1H, J = 6.6 Hz), 3.45 - 3.33 (m, 4H), 2.53 (bm, 4H)2H), 2.42-2.35 (m, 2H), 1.47 (d, 3H, J = 6.6 Hz), 1.44 (s, 9H). ¹³C NMR (75 MHz, CDCl₃): δ 154.7, 140.1, 134.0, 131.5, 128.7, 127.4, 125.5, 125.3, 125.3, 124.6, 124.0, 79.4, 61.5, 50.5, 43.8, 28.4. 18.7.

1-[1-(1-Naphthyl)ethyl]-4-(4-methoxybenzylamino)carbonylpiperazine (20). To a solution of Boc 18 (32 mg, 0.094 mmol) in dry CH₂Cl₂ (2 mL) was added trifluoroacetic acid (0.3 mL) at 0 °C. The mixture was allowed to stir for 1 h at 23 °C and then was concentrated under reduced pressure. To the residue was added toluene, and the mixture was concentrated under reduced pressure to give crude compound 19. To a solution of 1,1'-carbonyldiimidazole (18.6 mg, 0.11 mmol) in dry CH₂Cl₂ (2 mL) was added dropwise 4-methoxybenzylamine 5c (15.7 µL, 0.12 mmol) at 0 °C under argon atmosphere, and the mixture was allowed to stir for 4 h at 23 °C. The mixture was added dropwise to a solution of 19 in dry CH₂Cl₂ (1 mL). The mixture was allowed to stir for 24 h at 23 °C and then was concentrated under reduced pressure. The residue was purified by silica gel column chromatography to furnish compound **20** (34 mg, 90%) as a white amorphous solid, $R_f = 0.57 (CH_2Cl_2/$ MeOH = 9:1). ¹H NMR (400 MHz, CDCl₃): δ 8.32 (d, 1H, J = 7.5 Hz), 7.81-7.78 (m, 1H), 7.69 (d, 1H, J = 8.1 Hz), 7.52 (d, 1H, J = 7.1 Hz), 7.44-7.36 (m, 3H), 7.13 (d, 2H, J = 8.6 Hz), 6.78 (d, 2H, J = 8.6 Hz), 4.22 (s, 2H), 4.07 (q, 1H, J = 6.6 Hz),3.72 (s, 3H), 3.34–3.21 (m, 4H), 2.55–2.50 (m, 2H), 2.39–2.34 (m, 2H), 1.42 (d, 3H, J = 6.6 Hz). ¹³C NMR (100 MHz, CDCl₃): δ 160.1, 160.0, 159.4, 141.1, 135.4, 132.9, 130.2, 130.1, 130.0, 128.9, 127.0, 126.8, 126.0, 125.2, 115.2, 62.6, 56.6, 51.7, 45.4, 45.2, 20.0. IR (neat): 3340, 2925, 1618, 1512, 1248 cm⁻ MS (ESI): m/z 404 [M + H]⁺. HRMS (ESI), calcd for C₂₅H₃₀-N₃O₂ 404.2338, found 404.2336.

Molecular Modeling. Computational analyses utilized the Sybyl 8.1 suite (Tripos, Inc.) and the GOLD docking program (CCDC) following previously described protocols.¹⁰

X-ray Crystallography. The complex of inhibitor 15g with purified PLpro was formed prior to crystallization by incubating 10 mg/mL PLpro (in 20 mM Tris, pH 7.5, 10 mM DTT) with 2 mM 15g at 4 °C for 16 h. Diffraction-quality crystals grew from a sitting drop containing 5 mg/mL PLpro, 1 mM 15g, 1 M (NH₄)₂SO₄, 50 mM MES, pH 6.5, and 2.5% PEG 400. Crystals were flash-frozen in liquid nitrogen and then transferred into a dry nitrogen stream at 100 K for X-ray data collection. The data set of the complex was collected at the Southeast Regional Collaborative Access Team (SER-CAT) beamline at the Advanced Photon Source, Argonne National Laboratory. Data were processed and scaled using the HKL2000 program suite. Crystals belonged to the space group C_2 , with two monomers in the asymmetric unit. The inhibitor-complexed structure was solved to 2.63 Å by molecular replacement using the SARS-CoV PLpro apoenzyme structure (PDB entry 2FE8) as a search model in the AMoRe program of the CCP4 suite. Manual model building was performed using Wincoot, and iterative rounds of positional and *B*-factor refinement and map building were performed using CNS. The structure was deposited under PDB code 3MJ5.

SARS-CoV Antiviral and PLpro Inhibition Assays. SARS-CoV antiviral assays and PLpro inhibition assays were performed as previously described.¹⁰

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Supporting Information Available: HPLC and HRMS data of inhibitors. This material is available free of charge via the Internet at http://pubs.acs.org.

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